

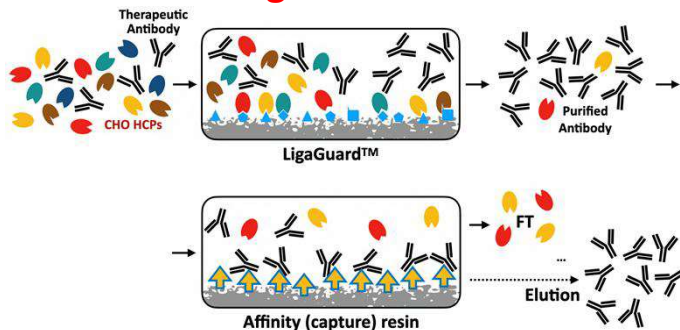
# Clearance of host cell proteins from CHO cell culture harvests via “Flow-through affinity chromatography” using LigaGuard™ resins

Sobhana Sripada, Wenning Chu, Stefano Menegatti, Jae Sly, Michael Crapazano

## - Introduction -

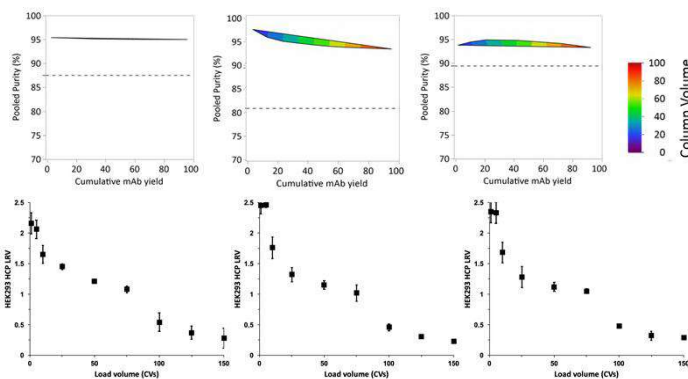
- Purification of therapeutic monoclonal antibodies (mAbs) consists of the removal of product-related (aggregates, fragments, charged variants, etc.) and process-related (host cell proteins (HCPs) and DNA etc.) impurities;
- Chinese Hamster Ovary harvested cell culture fluids (CHO HCCFs) contain, together with the mAb product, a substantial amount of HCPs (up to 0.5 mg/mL), which are diverse in titer, biophysical properties, and safety profile;
- HCP clearance poses major challenges due to the ability of highly immunogenic HCPs to associate with and/or degrade the mAb product;
- These “high-risk HCPs” impose taxing optimization of both Protein A purification and polishing;

## - LigaGuard™ -

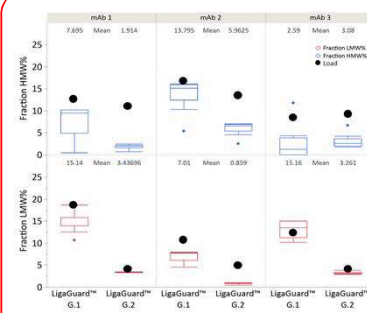


- Our work demonstrates that LigaGuard resin has the potential to:
  - **De-risk biomanufacturing:** by effectively clearing all HCPs, including the HR-HCPs that commercial resins may struggle to remove, LigaGuard resin provides a significant contribution towards safeguarding the stability and safety of special mAb products;
  - **Increase process flexibility and robustness** by widening the "space of design" where high mAb yield and purity are achieved and minimizing the impact of variability of HCP titer and composition in the harvest;
  - **Reduce CAPEX and OPEX** by enabling an agile purification sequence fully operated in flow-through mode, which are prized for their compact footprint and reduced volume of aqueous service streams;
  - **Enable purification processes of emerging therapeutic modalities**, such as gene therapy and cell therapies.

## - Results -



- Static CHP HCP binding capacity of LigaGuard resin > 20 g HCP per L resin;
- LigaGuard resin affords mAb yield of 93 - 96% irrespective of HCCF properties;
- HCP clearance (logarithmic removal value, LRV):
  - Average LRV > 1.5 for loading up to 30 CVs;
  - Average LRV ~ 1.3 – 1.4 for loading up to 75 CVs;
  - Average LRV ~ 1.1 – 1.2 for loading up to 100 CVs;



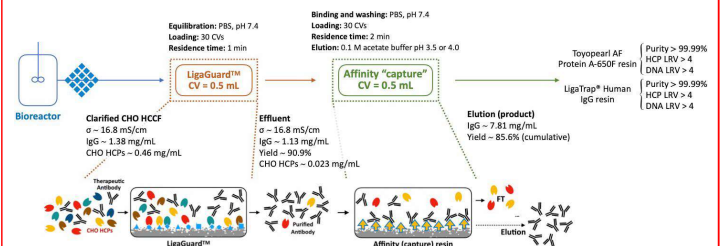
- Analysis of LigaGuard effluents via SEC HPLC demonstrated the selective recovery of mAb monomers, while mAb aggregates are cleared; we hypothesize that the capture of HCPs favors the removal of HCP-associated mAb aggregates.



- HCPs in the effluents were identified via proteomics analysis.
- Statistical analysis identified **significantly removed HCPs** in comparison with corresponding CHO HCCF feedstocks.
- Different peptides in LigaGuard removed specific subsets of HCPs in certain fractions.
- >95% of HCPs identified in the CHO HCCF were effectively cleared across all tested feedstocks using G.2 LigaGuard

- **Effective clearance of high-risk HCPs** was documented, including phospholipase B-like 2 protein (PLBL-2), Heat shock protein 70 (HSP70), Cathepsins A, B, D, Z, serine proteases, carboxypeptidases, glutathione transferases, metalloproteases, etc.
- **LigaGuard resin can be regenerated**, making it amenable to both single-use or multiple use formats.

## - LigaGuard™-based Process -



- **Documented removal of Chromatin-based and HCP-laden aggregates** prior to the affinity capture column, thus reducing HCP persistence;
- **Two-Step HCP LRV > 4 and DNA LRV > 4;**
- Improved Protein A performance (elution yield and lifetime);
- Improved stability of the mAb product (long-term colloidal stability of the mAb under cold storage).

## - Publications -

- S. Sripada, [...], R.G. Carbonell, A.M. Lenhoff, S.M. Cramer, J. Bill, D. Roush, and S. Menegatti. "Towards continuous mAb purification: clearance of host cell proteins from CHO cell culture harvests via “flow-through affinity chromatography” using peptide-based adsorbents." *Biotechnology and Bioengineering* (2022).
- A. R. Lavoie, [...], R.G. Carbonell, and S. Menegatti. "Removal of host cell proteins from cell culture fluids by weak partitioning chromatography using peptide-based adsorbents." *Separation and Purification Technology* 257 (2021): 117890.
- A. R. Lavoie, [...], R.G. Carbonell, and S. Menegatti. "Targeted capture of Chinese hamster ovary host cell proteins: Peptide ligand binding by proteomic analysis." *Biotechnology and Bioengineering* 117, no. 2 (2020): 438-452.
- A. R. Lavoie, [...], R.G. Carbonell, and S. Menegatti. "Targeted capture of Chinese hamster ovary host cell proteins: peptide ligand discovery." *International journal of molecular sciences* 20, no. 7 (2019): 1729.
- Peptide ligands for capture of host cell proteins (US20220009959A1).